



## Tema -6

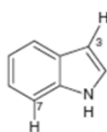
### Preparing the Sample



## Sample



A correct sample preparation allows obtained the spectra with a good line shape



- The five-bond coupling (H3-H7) are very small – similar to the natural line width of a typical NMR signal.
- Only in the best conditions are possible to observe this couplings (differences about one part in  $10^9$ )
- It is necessary that the magnetic field is very homogeneous, so that all molecules experience the same field regardless of position in the NMR tube

### How Much Sample Do I need ?

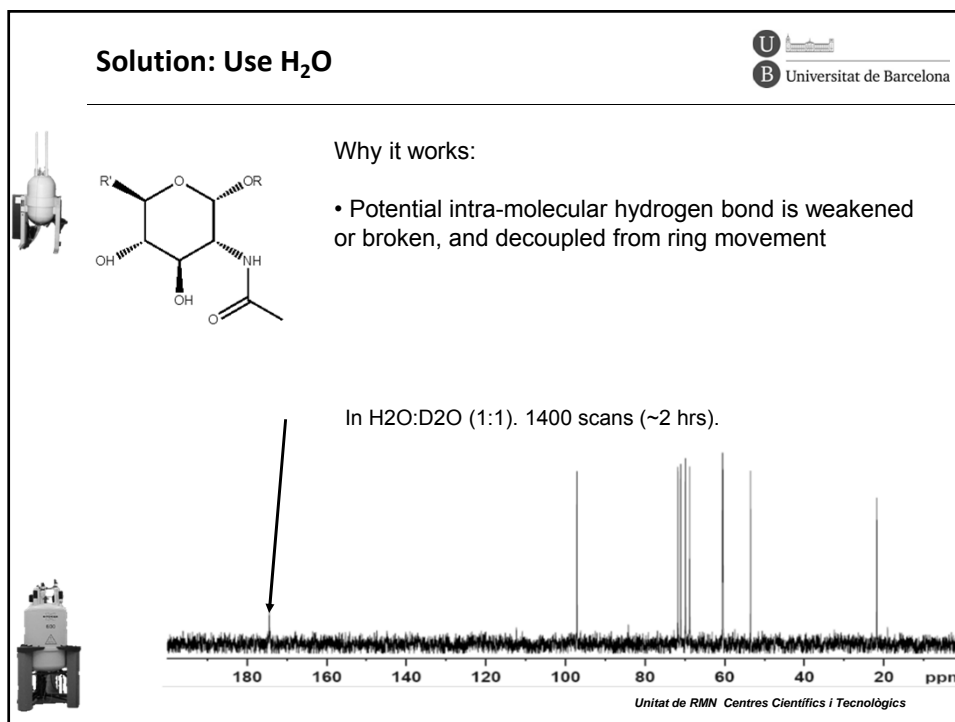
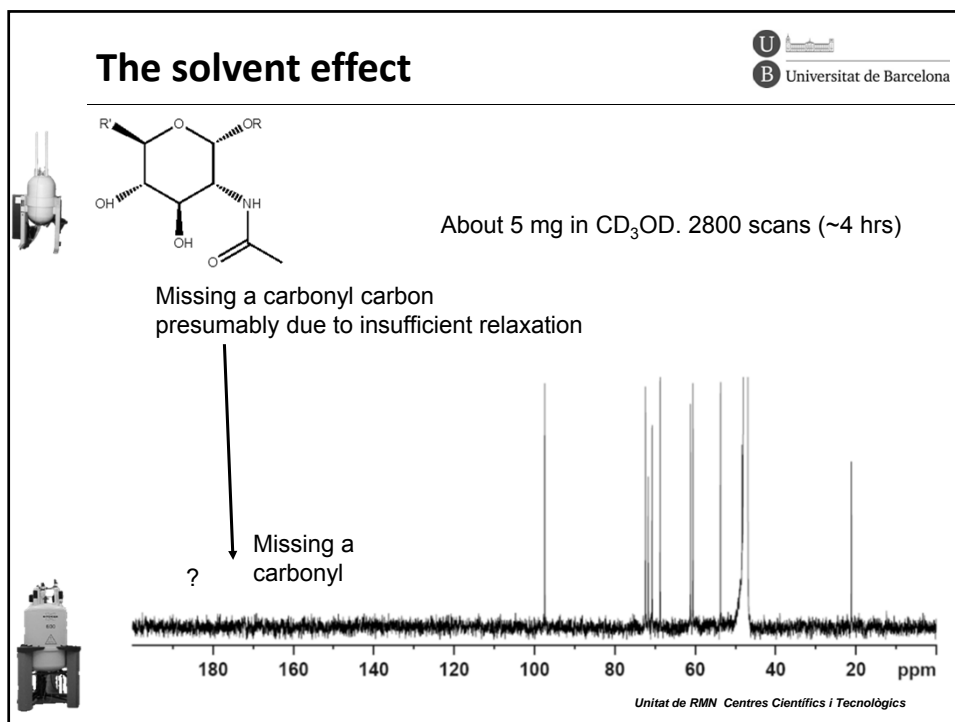
- Nucleus to observe
- Probe
- Magnetic Field
- How pure is the sample ?
- Molecular weight
- What experiment you need to perform?

Started point

Field	$^1\text{H}$	$^{13}\text{C}$
400	10-5 mg	30-50 mg
500	5-2 mg	20-10 mg
600	2-1 mg	10-5 mg



**If your spectra has a high noise level after 5 minutes, probably you need more than 1h 20 min (increase the signal by factor of four)**



## Solvent selection



Whenever possible you should use deuterated solvents

Exceptions: peptides, proteins and .....



**Selected solvent must dissolve completely the sample**  
**If the sample is not the solution, then it is invisible to the NMR spectrometer. Only degrades the homogeneity**

### Deutero Cloroform

- The most useful and cheaper solvent. It can dissolve a variety of compounds (polar and nonpolar)
- The residual CHCl<sub>3</sub> at chemical shift 7.26 ppm (or 7.24).
- You can add a drops to DMSO-d<sub>6</sub> but the chemical shift of CHCl<sub>3</sub> change,
- It should be avoided in samples with salts. As aprotic, that does not facilitate fast transfer of exchangeable (alcohols, amines ...) You can see the different species or broad signals.

The CDCl<sub>3</sub> can be acidic, and acid sensitive compounds may decompose when dissolved in CDCl<sub>3</sub>.



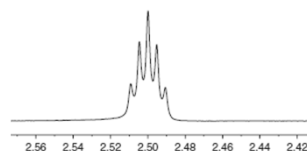
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## DMSO-d<sub>6</sub>



### Deutero Dimethyl Sulfoxide (D<sub>6</sub>-DMSO)

- Deutero dimethyl sulfoxide (D<sub>6</sub>-DMSO). Can dissolve a wide variety of compounds, including relatively insoluble heterocyclic compounds and salts.
- Can be a good choice for high temperature work – it could be taken up to above 140°C.
- Allows the observation exchangeable protons



### Drawbacks

- It's relatively viscous, and this causes some degree of line-broadening.
- The dms is freezing at 18.5°C
- The worst problem with DMSO, is its affinity for water. The the labile deuterons (and protons) of water (4.06 ppm) are available to exchange with labile protons in the chemist's sample and can result in inaccurate integration ratios.
- Extreme care should be taken when handling DMSO solutions, as one of its other characteristics is its ability to absorb through the skin taking your sample with it!



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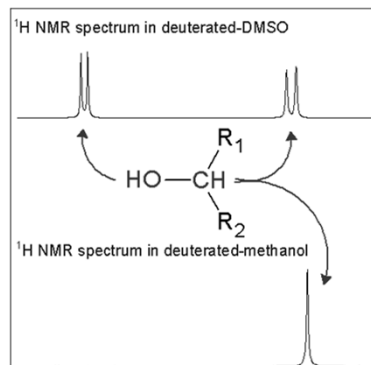
## Other Solvents



### Deutero Methanol (CD3OD)

This is a very polar solvent, suitable for salts and extremely polar compounds. It has a very high affinity for water and is almost impossible to keep dry. Its water peak is sharper at around 4.8 ppm. The residual CD<sub>2</sub>HOD is observed at 3.3 ppm.

Its main disadvantage is that it will exchange ionizable protons in your sample for deuterons, and hence they will be lost from the spectrum, e.g., -OH, -NH and even -CONH<sub>2</sub>, though these can often be relatively slow to exchange. Also, protons  $\alpha$  to carbonyl groups may exchange through the enol mechanism.



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## More solvents



### Deutero Water (D<sub>2</sub>O)

- D<sub>2</sub>O is even more polar than D<sub>4</sub>-methanol and rather limited in its use for that reason usually for salts only.
- It exchanges all acidic protons readily and exhibits a strong HOD signal at about 4.9 ppm.
- Sometimes it can be used 1:9 mixtures D<sub>2</sub>O/H<sub>2</sub>O to observe amide protons

### More solvents

Trifluoroacetic Acid (CF<sub>3</sub>COOH)

Pyridine- d<sub>5</sub>

C<sub>6</sub>D<sub>6</sub> or CCl<sub>4</sub>

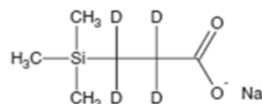
**Not recommended (carcinogenic)**

### Reference

TMS (TetraMethyl Silane)

TSP (3-(Trimethylsilyl) propionic-2,2,3,3-D<sub>4</sub> acid)

Residual solvent signals



*J. Org. Chem.* **1997**, *62*, 7512-7515. "NMR Chemical Shifts of Common Laboratory Solvents as Trace Impurities"

*Organometallics* **2010**, *29*, 2176-2179. "NMR Chemical Shifts of Trace Impurities: Common Laboratory Solvents, Organics, and Gases in Deuterated Solvents Relevant to the Organometallic Chemist"

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## The most common solvents



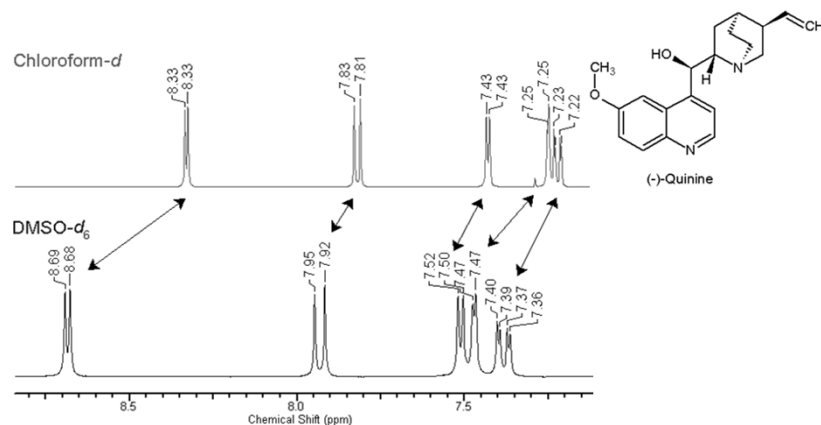
[http://www.isotope.com/uploads/File/NMR\\_Solvent\\_Data\\_Chart.pdf](http://www.isotope.com/uploads/File/NMR_Solvent_Data_Chart.pdf)

Solvent	Formula	H-NMR shift	C-NMR shift	multiplet	$J_{C,H}$ (Hz)	mp (°C)	bp(°C)	comments
Chloroform-d	$CDCl_3$	7.24	77.0	triplet	32	-64	61	
Acetone-d <sub>6</sub>	$CD_3COCD_3$	2.04	29.8	septet	20	-95	56	
			206.3	multiplet	<1			appears as singlet mostly
Benzene-d <sub>6</sub>	$C_6D_6$	7.2	128.0	triplet	24	6	80	carcinogen
Acetonitrile-d <sub>3</sub>	$CD_3CN$	1.93	1.3	septet	21	-45	82	
			117.7	multiplet	<1			appears as singlet mostly
Dichloromethane-d <sub>2</sub>	$CD_2Cl_2$	5.32	53.5	quintet	21	-97	40	
Dimethylsulfoxide-d <sub>6</sub>	$CD_3SOCD_3$	2.49	39.7	septet	21	19	189	
Methanol-d <sub>4</sub>	$CD_3OD$	3.35, 4.78	49.3	septet	21	-98	64	
Tetrahydrofuran-d <sub>8</sub>	$C_4D_8O$	1.73, 3.58	25.5	quintet	21	-108	66	
			67.7	quintet	22			
Toluene-d <sub>8</sub>	$C_7D_8$	2.30, 7.19				-95	111	
Pyridine-d <sub>5</sub>	$C_5D_5N$	7.19, 7.55, 8.71	123.5	triplet	25	-42	115	
			135.5	triplet	24			
			149.5	triplet	27			
Water-d <sub>2</sub>	$D_2O$	4.65				0	100	
Acetic acid-d <sub>4</sub>	$CD_3COOD$	2.03, 11.53	20.0	septet	20	17	118	
			178.4	multiplet	<1			appears as singlet mostly
Trifluoroacetic acid-d	$CF_3COOD$	11.5	116.5	quartet	283	-15	72	C-F-coupling
			164.4	quartet	44			C-F-coupling
Dioxane-d <sub>8</sub>	$C_6D_{12}O_2$	3.58	66.5	quintet	22	12	102	
Tetramethylsilane (TMS)	$Si(CH_3)_4$	0.00	0.00	singlet				internal reference

## Effect of change of solvent




Certain experimental conditions, such as varying the temperature or using different solvents, may cause changes in chemical shifts  
When it comes to referencing 2D NMR data, use the 1D NMR data collected at the same temperature and solvent.

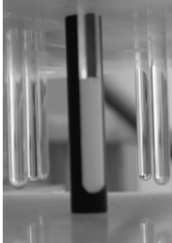





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## The Sample


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- Adjust the sample concentration to the solubility  
Avoid product precipitation
- Use a single deuterated solvent  
Reference for lock
- Avoid heterogeneous samples.  
Avoid air bubbles, suspended particles, sample separation
- The low quality NMR tubes → distorts magnetic field homogeneity  
Breaks easily → damage the NMR probe
- Adjust the Properly position NMR sample in the magnet with the NMR gauge

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## NMR Sample Preparation

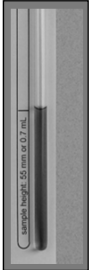
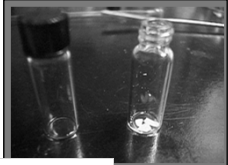

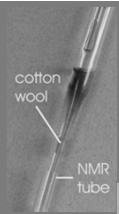
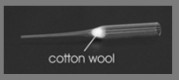


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For  $^1\text{H}$  Use about 5-25 mg of product

1-0.75 ml deuterated solvent  
The sample should be about 4.5-5.5 cm of liquid

The sample must be free of suspended material: You can filtering using a cotton wool

labeled your sample

NO

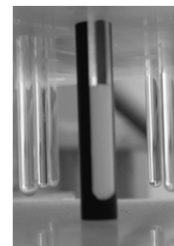
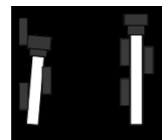
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## The quality of the spectra is not enough?



### • Failure due to sample preparation

- Dilute samples do not benefit from a short liquid length. The sample should be about 4.5-5.5 cm of liquid
- The height of the NMR-tube in the spinner is not correct
- Contaminations affecting spectral quality. You should avoid:
  - Paramagnetic Substances
  - High salt concentration
  - Particles "fishes" and not dissolved compounds
  - Mixes of deuterated solvents



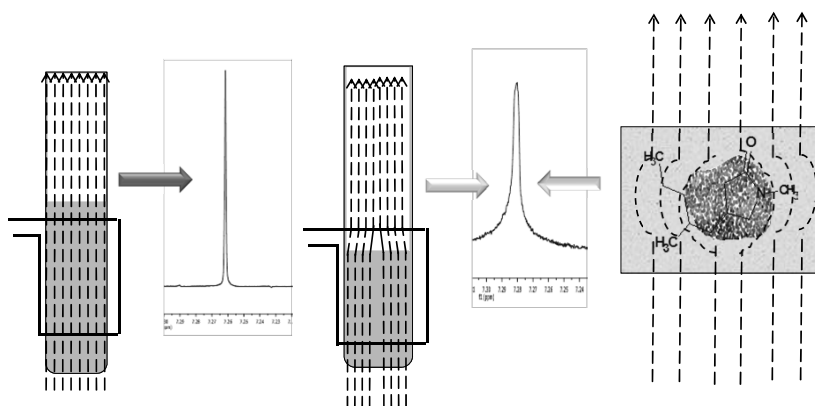
The time required for shimming increases significantly and may be impossible to obtain a quality spectra

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## Effects due to incorrect sample preparation



An insufficient volume or the presence of precipitate in the solution leads to a distortion of magnetic field lines. The result is a change in the line shape in the signals of the spectrum or a peak splitting



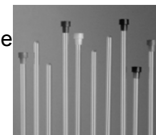
Overfilling the NMR tubes may cause problems due to temperature gradients in the sample

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## NMR Sample Tube Care of tube and cleaning



- Do not heat the NMR tube in an oven, the tube will warp and may cause probe damage. Bent tubes may cause severe probe damage.
- Clean the NMR tube with a suitable solvent and dry with acetone and the nitrogen or air
- Tube caps are very cheap and disposable, dirty caps or old caps can contaminate your sample
- Use only EIGHT-INCH tubes on the SMS sample changer!
- The price of NMR tubes range from <math><2-3\text{€}</math> a piece to >50€. For high-quality NOE-based spectra at higher fields use high-quality tubes



**Recommended for 400-500 MHz:  
Wilmad ref PP 507 or PP 528 (7-14 \$) Length: 8 inches**

[http://www.wilmad-labglass.com/services/technical\\_NMR\\_EPR.jsp](http://www.wilmad-labglass.com/services/technical_NMR_EPR.jsp)

<http://www.cortecnet.com>

<http://www.newera-spectro.com/>



**Wilmad**

[http://www.wilmad-labglass.com/services/NMR\\_001.jsp](http://www.wilmad-labglass.com/services/NMR_001.jsp)

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